

# Diabetes and Alzheimer's Disease - Is There a Connection?

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## Abstract

It has been known for some time that diabetes may be associated with impaired cognitive function. During the last decade, epidemiological data have emerged suggesting a linkage between diabetes, particularly type 2 diabetes, and Alzheimer's disease (AD). There is evidence to suggest that impaired activities of neurotrophic factors such as insulin, IGF-1 and NGF, which occur in both diabetes and AD, may provide a mechanistic link between the two disorders. An additional probable factor that has been less evaluated to date is hypercholesterolemia, a common accompaniment to type 2 diabetes. Increased cholesterol availability is believed to play a crucial role in the abnormal metabolism of amyloid precursor protein leading to accumulation of amyloid-β. Im-

#### Introduction

C ognitive impairments are more common in diabetic patients than in non-diabetic subjects [1-5], which is in part due to ischemic events resulting from cerebral micro- and/or macrovascular disease or to repeated episodes of severe hypoglycemia. These conditions have been referred to as secondary diabetic encephalopathy [6]. However, during the last decade, there is accumulating evidence suggesting that cognitive dysfunction is also caused by diabetic dysmetabolism, so-called primary diabetic encephalopathy [2, 3, 5, 6].

This appears to be true also in experimental models of diabetes. In streptozotocin-induced diabetic rats, impaired cognitive performances have been associated with impaired hippocampal plasticity, changes which paired insulin signaling in particular appears to be involved in hyperphosphorylation of the tau protein, which constitutes neurofibrillary tangles in AD. The linkage between abnormal amyloid metabolism and phosphor-tau is likely to be provided by the activation of caspases both by increased amyloid- $\beta$  and by impaired insulin signaling. Although the details of many of these components still await evaluation, it appears clear that commonalities exist in the underlying pathogenesis of diabetes and Alzheimer's disease. In this review we provide a brief update on linkages between these two diverse but common disorders.

**Keywords**: diabetes · Alzheimer's disease · insulin · hypercholesterolemia · dementia

are reversed by insulin treatment [7]. In type 1 diabetic BB/Wor rats, progressively impaired cognitive function is associated with suppressed insulin and insulinlike growth factor I (IGF-1) actions and neuronal apoptosis in hippocampus [8], changes which are significantly prevented by insulinomimetic C-peptide [9, 10].

The increased risk for cognitive dysfunction affects both type 1 and type 2 diabetic patients [1-5], suggesting that hyperglycemia or altered insulin signaling transduction, or both, are involved. More relevant deficits, however, occur in patients with type 2 diabetes in whom there is an increased risk for developing Alzheimer's disease (AD) [11, 12], suggesting that additional factors may be involved. In addition to hyperglycemia and impaired insulin action, type 2 diabetes is commonly associated with hypercholesterolemia, hy-

perlipidemia and hypertension, which may provide additional risk factors. In addition, aging alone is probably a modulating factor. Experimental studies have demonstrated significantly more severe abnormalities in the expression of amyloid precursor protein (APP),  $\beta$ -secretase, amyloid- $\beta$  (A $\beta$ ) and phosphorylated tau in the type 2 BBZDR/Wor rat model as compared to its type 1 counterpart, the BB/Wor rat [13]. The type 2 diabetic model is characterized by insulin resistant hyperglycemia, hypercholesterolemia, hyperlipidemia and hypertension, hence closely replicating the common clinical picture of type 2 human diabetes [14]. The combination of these factors comprises the "metabolic syndrome." Some of these factors have been identified as independent predictors of cerebrovascular disease, accelerated cognitive dysfunction and dementia [15, 16]. Therefore, the clustering of several potential pathogenetic factors may interact mechanistically at various levels in type 2 diabetes to produce the basis

tural substrates of Alzheimer's disease [3, 5]. Recent research has implicated hyperglycemia, insulin-resistance and impaired insulin and insulin-like growth factor-1 (IGF-1) signaling with activation of so-called stress (or tau) kinases as mechanisms in the production of phosphorylated tau, a characteristic hallmark of AD [17-19]. Less attention has been devoted to the possible role of hypercholesterolemia and its role in APP metabolism, abnormal A $\beta$  handling and deposition, the second characteristic hallmark of AD.

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In this review we explore the mechanistic linkages between APP metabolism, tau abnormalities and the role of cholesterol metabolism in AD and how these abnormalities can be linked to diabetes.

## **Insulin effects**

The emerging cognitive impairment and even the possible relationship with AD in diabetes are supported by epidemiological studies [1-4], as well as findings in animal models of diabetes [6-10,20-22]. Experimental data have demonstrated increased expression of APP,  $\beta$ -secretase, A $\beta$ 42 and hyperphorylated tau in hippocampi and frontal cortex of BB rats. Hence, taken together, these findings strongly suggest mechanistic and sequential links between diabetes, impaired cognitive function and molecular and structural AD-like changes.

In humans, aging alone is associated with decreased metabolic turnover, decreased glucose utilization as well as declining insulin and IGF-1 signal transduction due to receptor desensitization [23-25]. These abnormalities are magnified in Alzheimer's disease, with degradation of both the insulin and IGF-1 receptor and their consequent effects on glucose metabolism [26-28]. Other factors that contribute indirectly to impaired insulin signaling include cortisol, catecholamines and cholesterol via the caveolin signaling pathway (see below).

Furthermore, the APP derivatives Aβ40 and Aβ42 reduce the binding of insulin to its receptor, either through binding or via an ATP-mediated disruption of autophosphorylation [29, 30], hence creating a potentially self-perpetuating mechanism. Other mechanisms caused by the convergence of insulin and extracellular A $\beta$  are the competition for insulin degrading enzyme (IDE) [31]. Due to counterregulation by GLUT transporters (GLUT1 and 3) at the blood-brain barrier [32], it appears that CNS insulin levels are increased in systematically insulinopenic situations. Hence, elevated CNS insulin levels will consume more of the already reduced levels of IDE [33], thereby intervening with the degradation of and promoting the extracellular deposition of Aβ42 [31]. Therefore, perturbations in insulin levels and its signal transduction activity will potentially provide several sites of action in promoting AD-like pathologies.

## Effects of cholesterol

Epidemiological evidence exists linking elevated plasma cholesterol and lipoprotein levels with AD development [15, 16, 34]. Additionally, patients taking cholesterol-lowering drugs have been found to have a lower incidence of AD [35], although some studies have proven to be inconclusive [36]. Experimental studies examining the loading or depletion of cholesterol both in vivo [37-39] and in cell cultures [40] have demonstrated links between high cholesterol and increased A $\beta$  production; however, the mechanisms underlying such linkages are poorly understood. It should be mentioned, though, that brain cholesterol is mostly independent of dietary uptake or hepatic synthesis but appears to be mainly derived from in situ synthesis [41] (Figure 1). There is evidence to suggest that statins, hydroxyl-3-methylglutaryl-CoA inhibitors, not only lower cholesterol levels (systemic and endogenous) but also suppress  $\beta$ -secretase activity in lipid rafts and increase  $\alpha$ -secretase, thereby directly effecting APP metabolism [42].

The expression of apolipoprotein E allele 4e (Apo4E) is a major risk factor for the development of sporadic Alzheimer's disease. It is a lipoprotein that carries and facilitates the transport and incorporation



Figure 1. Simplified flow-chart of interactions between high systemic and endogenous cholesterol levels and insulin resistance in nerve cells. Shown are effects on amyloidogenic and non-amyloidogenic APP metabolism (blue) as well as intra- and extracellular accumulation of AB products. The insulin receptor (IR) is located within the caveolae and its signaling is promoted by caveolin-1, whereas insulin resistance (red), as existent in type 2 diabetes or modeled by Wortmannin (yellow), interferes with insulin signaling. Impaired insulin signaling inhibits the release of sAPPa from the intra- to the extracellular space (blue). It also promotes the activation of caspases and via the activation of so-called stress kinases enhances the phosphorylation of tau. Increased extra- and intracellular cholesterol becomes enriched in lipid rafts, where it is coupled with up-regulation of APP as well as  $\alpha$ - and  $\beta$ secretase resulting in increased accumulation of A $\beta$  and CTFs (amyloidogenic pathway, blue). These amyloid peptides and fragments are released to the extracellular space or remain intracellular. Aß products activate caspases, which in turn promote the splicing of tau and subsequent phosphorylation by stress kinases. Apo4E enhances the incorporation of cholesterol into lipid rafts, promotes formation of Aß products, interferes with insulin signaling and inhibits non-amyloidogenic formation of sAPPa. For additional metabolic mechanisms see the text.  $\rightarrow$  indicates promoting effects, - indicates inhibitory effects.

of cholesterol within lipid rafts in caveolae [43] (Figure 1). It is associated with caveolae, increasing the formation of A $\beta$  fibrils and decreasing soluble amyloid precursor protein alpha (sAPP $\alpha$ ), to yield a reciprocal regulation on A $\beta$  and sAPP $\alpha$  [44]. Apo 4E inhibits intermediaries of signal transduction pathways that lead to phosphorylation of PI, which is impaired in AD [45] (Figure 1). This results subsequently in intracellular Ca<sup>++</sup> release and perturbation of PI3 and DAG [46] leading to increased apoptotic stress. Increased cholesterol levels are associated with an increased number of caveolae, harboring lipid rafts in which cholesterol, tometabolism underlies the formation of amyloid plaques, one of the characteristic hallmarks of AD. Abnormal APP processing is believed to play a central and probably initiating role in AD and gives rise to A $\beta$  (Figure 1). APP is a transmembranous protein, the physiological role of which remains unknown, but may be involved in neuronal regeneration. It is metabolized along two pathways; an amyloidogenic and a non-amyloidogenic pathway [42, 43]. In the latter it is cleaved by  $\alpha$ -secretase producing sAPP $\alpha$  but not A $\beta$ . In the former it is cleaved by  $\beta$ secretase and  $\beta$ APP is produced, which is further cleaved by  $\gamma$ -secretase, ultimately yielding A $\beta$ , most

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Lipid rafts are thought

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tors are concentrated

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signaling pathways are

domain of calveolin-1

[48], with the excep-

tion of insulin signaling, which is enhanced

by calveolin-1 [29, 30, 49] (Figure 1). Choles-

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There is growing evidence that amyloidogenic APP processing is facilitated in cholesterol-rich lipid rafts, whereas non-amyloidogenic APP processing by  $\alpha$ secretase may occur outside the rafts [37], resulting in sAPP $\alpha$ , which does not produce A $\beta$  (Figure 1). The production of sAPPa is downregulated under high cholesterol conditions, whereas depletion of cellular cholesterol increases the levels of sAPP $\alpha$  [52]. Amyloidogenic processing of APP occurs in lipid rafts where  $\beta$ - and  $\gamma$ -secretases are found as APP, C-terminal fragments (CTFs) and A $\beta$  itself [53, 54]. It is believed that APP is localized in two separate pools, one associated with lipid rafts and the other with non-raft domains, and would be accessible through  $\alpha$ -,  $\beta$ - and  $\gamma$ secretase (Figure 1). With respect to  $\beta$ -secretase, it appears to act at the plasma membrane level, as well as in ER and in the trans-Golgi network. The subsequent A $\beta$  formation at the plasma membrane and CTFs are believed to be secreted, whereas those originating from the ER and trans-Golgi network probably remain intracellular [55] (Figure 1).

Further mechanisms that are likely to involve abnormal APP metabolism relate to impaired insulin signaling that has been demonstrated in the frontal cortex of the BBZDR/Wor rat [7]. Deficiencies in insulin, IGF-1 and NGF converge in their signal transduction, diminishing tyrosine-kinase signaling and impairing PI3 kinase activity, thereby activating stress kinases such as JNK and p38 which lead to activation of caspase 3. For instance, inhibition of insulin signaling by Wortmannin hampers the release of sAPP $\alpha$  and A $\beta$ from the intra- to the extracellular compartment [44, 56] (Figure 1). Insulin deficiency activates FAS including NGFR-p75 with activation of caspase 8, which has been implicated in the splicing of APP into CTFs and into intracellular processing into AB42 [57]. Oxidative stress and mitochondrial dysfunction with activation of caspase 9 have been invoked in abnormal APP processing [46, 47]. Therefore, several apoptotic stressors are likely to contribute to abnormal intracellular processing of APP.

### Apoptotic mechanisms

Insulin, C-peptide and IGF-1 all exert antiapoptotic functions [10, 58, 59]. In diabetic neuronal tissues activation of a variety of apoptotic pathways has been identified. Deficiencies of insulin, C-peptide, IGF-1 and NGF converge in their signal transduction diminishing tyrosine-kinase signaling and impaired PI3 kinase activity, thereby activating stress kinases-like JNK and p38, particularly JNK, which lead to the activation of caspase 3 [58].

Insulin and C-peptide deficiencies activate the FAS receptor family as well as NGFR-p75, which has been implicated in apoptosis. FAS activation leads via FADD to the activation of caspase 8 or caspaseindependent apoptosis [58, 59]. Caspase 8 has been implied in the C-terminal splicing of APP to CTFs and further processing of intracellular APP into soluble A $\beta$ 42 [60, 61] by increased activity of  $\beta$ -secretase [62]. Oxidative stress is also likely to contribute to activation of caspase 2 and 8 triggered by cytochrome C and AIF through induction of death receptors [63]. An apoptotic pathway invoked in the abnormal APP processing is caspase 9. Several stressors such as oxidative stress and mitochondrial dysfunction appear to be involved, as reflected by increased pro-apoptotic Bax, AIF and nuclear stainability for 8OHdG [22, 60, 61].

It therefore appears that several apoptotic stressors identified in diabetic brain [9, 10] may be involved in abnormal intracellular processing of APP forming both C-terminal fragments, as well as intracellular cleavage to A $\beta$ 40 and A $\beta$ 42 (Figure 1). The apoptotic stressors exert these functions before the cell undergoes apoptotic cell death, a phenomenon that appears to be regulated by upregulation of contraregulatory proteins, among others heat shock proteins 27 and 70, which have recently been demonstrated in diabetes [64, 65].

Decreased PI3-K activity and associated activation of stress-kinases such as JNK and p38 promote the abnormal processing of APP [66]. Phosphorylated p38 MAPK has been demonstrated in early Braak stages of AD. Indirect evidence for a mechanism for JNK and p38 kinases is provided by inhibition of insulin signaling by Wortmannin, which alters the metabolism of APP [67] and hampers the release of sAPP $\alpha$  and A $\beta$ from the intracellular to the extracellular compartment [68]. Under normal conditions, PKC and PKA promote utilization of non-amyloidogenic processing of APP by redistributing it to compartments of  $\alpha$ secretase activity with increase in sAPP $\alpha$  levels and decreased release of A $\beta$ , the so-called reciprocal relationship between these APP products [69].

Furthermore, JNK and p38 activations are involved in the hyperphosphorylation of tau, via A $\beta$ -induced oxidative stress and apoptosis. Puig *et al.* reported increased expression of JNK and p38 in brain homogenates and their immunocytochemical association with hyperphosphorylated tau in neurites surrounding amyloid plaques [70].

## Tau pathology

One of the most important pathobiological events in AD is the formation of hyperphosphorated tau, which leads to a toxic insult to terminal neurites and progressive retrograde neurite degeneration [71-73]. This is not specific for AD, but occurs in a variety of neurodegenerative conditions, the so-called tauopathies [74]. However, in these conditions, the tau pathology is not associated with amyloid pathology, suggesting that several mechanisms result in hyperphosphorylation of abnormal tau. The hyperphosphorylation of tau results in loss of its function in promoting the assembly and stabilization of microtubules. It also sequesters normal tau [75].

In AD,  $A\beta 42$  induces several apoptotic pathways [76-78]. Caspase activation is present in AD brains and active caspase is found within tangle-bearing neurons [79, 80]. Tau is initially cleaved by proteolytic caspases and undergoes subsequent hyperphosphorylation (Figure 1). The latter seems to be mainly mediated by increased GSK3ß activity, although decreased phosphatase activity of PP2B [81] and abnormal activity of the p25-Cdk5 complex [82] have been invoked. As referred to earlier, the insulin and related neurotrophic factor signal transduction activities are compromised in both AD and diabetes [6, 10, 28, 67], leading to inhibition of glucose metabolism, ATP formation and impaired PI3-kinase signaling. As a consequence, downstream PKB becomes downregulated, with disinhibition of GSK-3<sup>β</sup> and excessive phosphorylation of truncated tau [83-85] (Figure 1). Simultaneous disinhibition of GSK-3a may promote storage and misfolding of APP metabolites with secondary reduction of extracellular Aβ42 [86]. The subsequent accumulation

# References

- Biessels GJ, Kappelle AC, Bravenbaer B, Erkelens DW, Gispen WH. Cerebral function in diabetes mellitus. *Diabetolo*gia 1994. 37:643-650.
- Kramer L, Fasching P, Madl C, Schneider B, Damjancic P, Waldhausl W, Irsigler K, Grimm G. Previous episodes of hypoglycemic coma are not associated with permanent cognitive brain dysfunction in IDDM patients on intensive insulin treatment. *Diabetes* 1998. 47:1909-1914.
- Ott A, Stolk RP, van Harskamp F, Pols HA, Hofman A, Breteler MM. Diabetes mellitus and the risk of dementia: the Rotterdam study. *Neurology* 1999. 58:1937-1941.
- 4. Xu WL, Qui CX, Wahlin A, Winblad B, Fratiglioni L. Diabetes mellitus and risk of dementia in Kungsholmen project: a 6-year follow-up study. *Neurology* 2004. 63:1181-1186.
- Arvanitakis Z, Wilson RS, Bievias JL, Evans DA, Bennett DA. Diabetes mellitus and risk of Alzheimer's disease and decline in cognitive function. *Arch Neurol* 2004. 61:661-666.

of both A $\beta$ 42 and hyperphosphorylated tau leads to neurite and neuronal degeneration [71, 86, 87]. It is, therefore, unlikely that the commonalities between the sequence of molecular events leading to AD and the abnormalities of diabetic dysmetabolism are coincidental. Instead, evidence to date suggests causal relationships between these common disorders.

### Conclusions

There is no doubt that mechanistic linkages exist between diabetes and Alzheimer's disease. Although these are not completely defined, they provide today a very active area of investigative research. The major abnormalities that appear to present commonalities between the two entities are impaired neurotrophic actions, particularly by insulin, but probably also by IGF-1 and NGF. They result in increased apoptotic activities with abnormalities in the tau protein rendering it more susceptible to hyperphosphorylation. The abnormalities in insulin signaling provide several stress kinases, which facilitate excessive phosphorylation of tau. Increased exposure to cholesterol promotes abnormal APP metabolism with extra- and intracellular accumulation of toxic amyloid-ß products, which in turn activate several caspases promoting abnormal tau products. In these interactions hyperglycemia per se probably plays a lesser role. Both the accumulation of amyloid- $\beta$  products and hyperphosphorylated tau exert toxic effects on neuronal neurites with their subsequent degeneration and eventual neuronal death.

The further examination of the intricate and complex interplay between the pathogenetic events of these two disorders is likely to provide biologically meaningful targets for future therapy.

- Sima AAF, Kamiya H, Li ZG. Insulin, C-peptide hyperglycemia and central nervous system complications in diabetes. *Europ J Pharmacology* 2004. 490:187-197.
- Biessels GK, Kamel A, Urban IJ, Spruijt BM, Erkelens DW, Gispen WH. Water maze learning and hippocampal synaptic plasticity in streptozotocin-diabetic rats: effects of insulin treatment. *Brain Res* 1998. 800:125-135.
- Li ZG, Zhang W, Grunberger G, Sima AA. Hippocampal neuronal apoptosis in type 1 diabetes. *Brain Res* 2002. 946:212-231.
- Li ZG, Zhang W, Sima AA. C-peptide prevents hippocampal apoptosis in type 1 diabetes. Int J Exp Diab Res 2002. 3:241-246.
- Sima AA, Li ZG. The effect of C-peptide on cognitive dysfunction and hippocampal apoptosis in type 1 diabetes. *Diabetes* 2005. 54:1497-1505.
- 11. **Peila R, Rodriquez BL, Launer LJ**. Type 2 diabetes, APOE gene and the risk for dementia and related pathologies: The Honolulu-Asia Aging Study. *Diabetes* 2002. 51:1256-1262.

- Sinclair AJ, Girling AJ, Bayer AJ. Cognitive dysfunction in older subjects with diabetes mellitus: impact on diabetes selfmanagement and use of care services. All Wales Research into Elderly (AWARE) Study. *Diab Res Clin Prac* 2000. 50:203-212.
- 13. Li ZG, Zhang W, Sima AA. Increased beta-amyloid and phospho-tau in diabetic encephalopathy. *Diabetes* 2006. 55:A190.
- Sima AA, Zhang W, Xu G, Sugimoto K, Guberski D, Yorek MA. A comparison of diabetic polyneuropathy in type-2 diabetic BBZDR/Wor-rat and in type 1 diabetic BB/Worrat. *Diabetologia* 2000. 43:786-793.
- Kivipelto M, Helkala EL, Laakso MP, Hanninen T, Hallikainen M, Alhainen K, Soininen H, Tuomilehto J, Nissinen A. Midlife vascular risk factors and Alzheimer's disease later in life: longitudinal, population based study. *Brit Med* J 2001. 322:1447-1451.
- 16. Kivipelto M, Helkala EL, Laakso MP, Hanninen T, Hallikainen M, Alhainen K, Iivonen S, Mannermaa A, Tuomilehto J, Nissinen A, Soininen H. Apolipoprotein E epsilon 4 allele, elevated midlife total cholesterol level, and high midlife systolic blood pressure are independent risk factors for late-life Alzheimer disease. *Ann Intern Med* 2002. 137:149-155.
- Carro E, Torres-Aleman I. The role of insulin and insulinlike growth factor I in the molecular and cellular mechanisms underlying the pathology of Alzheimer's disease. *Eur J Pharm* 2004. 490:127-133.
- Steen E, Terry BM, Rivera EJ, Cannon JL, Neely TR, Tavares R, Xu XJ, Wands JR, de la Monte SM. Impaired insulin and insulin-like growth factor expression and signaling mechanisms in Alzheimer's disease - is this type 3 diabetes? J Alzheimer's Dis 2005. 7:63-80.
- Lambourne SL, Sellers LA, Bush TG, Choudhury SK, Emson PC, Suh YH, Wilkinson LS. Increased tau phosphorylation of mitogen-activated protein kinase consensus sites and cognitive decline in transgenic models for Alzheimer's disease and FTDP-17: evidence for distinct molecular processes underlying tau abnormalities. *Mol Cell Biol* 2005. 25:278-293.
- Flood JF, Mooradian AD, Morley JE. Characteristic of learning and memory in streptozotocin-induced diabetic mice. *Diabetes* 1990. 39:1391-1398.
- Kamal A, Biessels GJ, Duis SE, Gispen WH. Learning and hippocampal synaptic plasticity in streptozotocin-diabetic rats: interaction of diabetes and aging. *Diabetologia* 2000. 43:500-506.
- 22. Li ZG, Zhang W, Sima AA. The role of impaired insulin/IGF action in primary diabetic encephalopathy. *Brain Res* 2005. 1037:12-24.
- Messier C, Tsiakas M, Gagnon M, Desrockers A, Awad N. Effect of age and glucoregulation on cognitive performance. *Neurobiol Aging* 2003. 24:985-1003.
- Lindeboom J, Weinstein H. Neuropsychology of cognitive aging, minimal cognitive impairment, Alzheimer's disease, and vascular cognitive impairment. *Eur J Pharmacol* 2004. 490:83-86.
- Watson GS, Craft S. Modulation of memory by insulin and glucose: neuropsychological observations in Alzheimer's disease. *Eur J Pharmacol* 2004. 490:97-113.
- Vitiello MV, Merriam GR, Moe KE, Drolet G, Barsuess S, Kletke M, Schwarz RS. IGF-1 correlates with cognitive function in healthy older men and estrogenized women. *The Gerontologist* 1999. 38(S1):6.
- Gasparini L, Netzer WJ, Greengard P, Xu H. Does insulin dysfunction play a role in Alzheimer's disease? *Trends Pharmacol Sci* 2002. 23:288-293.

- Hoyer S. The aging brain. Changes in the neuronal insulin/insulin receptor signal transduction cascade trigger lateonset sporadic Alzheimer disease (SAD). A mini review. J Neurol Transm 2000. 109:991-1002.
- Xie L, Helmerhort E, Taddel K, Plewright B, van Bronswijk W, Martins R. Alzheimer's beta-amyloid peptides compete for insulin binding to the insulin receptor. J Neurosci 2002. 22:1-5.
- Chou CK, Dull TJ, Russel DS, Gherzi R, Lebwohl D, Ulrich A, Rosen OM. Human insulin receptors mutated at the ATP-binding site lack protein tyrosine kinase activity and fail to mediate post-receptor effects of insulin. J Biol Chem 1987. 262:1842-1847.
- 31. Farris W, Mansourian S, Chang Y, Lindsley L, Eckman EA, Frosch MP, Eckman CP, Tanzi RE, Selkoe DJ, Guenette S. Insulin degrading enzyme regulates the levels of insulin, amyloid beta-protein and the beta-amyloid precursor protein intracellular domain in vivo. *Proc Natl Acad Sci USA* 2003. 100:4162-4167.
- Dudli R, Kuschinsky W. Brain glucose transporters: relationship to local energy demands. *News Physiol Sci* 2001. 16:71-76.
- Cook DG, Leverenz JB, McMillan PJ. Reduced hippocampal insulin-degrading enzyme in late-onset Alzheimer's disease is associated with the apolipoprotein E-epsilon 4 allele. *Am J Pathol* 2003. 162:313-319.
- 34. Pappola MA, Bryant-Thomas TK, Herbert D, Pacheco J, Fabra Garcia M, Manjon M, Girones X, Henry TL, Matsubara E, Zambon D, et al. Mild hypercholesterolemia is an early risk factor for the development of Alzheimer amyloid pathology. *Neurology* 2003. 61:199-205.
- Wolozin B, Kellman W, Rousseau P, Celesia GG, Siegel G. Decreased prevalence of Alzheimer disease associated with 3-hydroxy-3-methyglutaryl coenzyme A reductase inhibitors. *Arch Neurol* 2000. 57:1439-1443.
- 36. Shepherd J, Blauw GJ, Murphy MB, Bollen EL, Buckley BM, Cobbe SM, Ford I, Gaw A, Hyland M, Jukema JW, et al. Pravastatin in elderly individuals at risk of vascular disease (PROSPER): a randomized controlled trial. *Lancet* 2002. 360:1623-1630.
- 37. Li L, Cao D, Garber DW, Kim H, Fukuchi K. Association of aortic atherosclerosis with cerebral beta-amyloidosis and learning deficits in a mouse model of Alzheimer's disease. *Am J Pathol* 2003. 163:2155-2164.
- Refolo LM, Malester B, LaFrancois J, Bryant-Thomas T, Wang R, Tint GS, Sambamurti K, Duff K, Pappolla MA. Hypercholesterolemia accelerates the Alzheimer's amyloid pathology in a transgenic mouse model. *Neurobiol Dis* 2000. 4:321-331.
- Shie FS, Jin LW, Cook DG, Leverenz JB, LeBoeuf RC. Diet induced hypercholesterolemia enhances brain A(beta) accumulation in transgenic mice. *Neuroreport* 2002. 25:455-459.
- 40. Fassbender K, Simons M, Bergmann C, Stroick M, Lutjohann D, Keller P, Runz H, Kuhl S, Bertsch T, von Bergmann K, Hennerici M, Beyreuther K, Hartmann T. Simvastatin strongly reduces levels of Alzheimer's disease betaamyloid peptides A(beta)42, A(beta)40 in vitro and in vivo. Proc Natl Acad Sci USA 2001. 98:5856-5861.
- 41. Jurevics H, Morell P. Cholesterol for synthesis of myelin is made locally, not imported into brain. J Neurochem 1995. 64:895-901.
- 42. Ehehalt R, Keller P, Haass C, Thiele C, Simons K. Amyloidogenic processing of Alzheimer beta-amyloid precursor

protein depends on lipid rafts. J Cell Biol 2003. 160:113-123.

- Sjogren M, Mielke M, Gustafson D, Zandi P, Skoog I. Cholesterol and Alzheimer's disease - is there a relation? *Mech Aging Develop* 2006. 126:138-147.
- Petanceska SS, Gandy S. The phosphatidylinositol 3-kinase inhibitor Wortmannin alters the metabolism of the Alzheimer's amyloid precursor protein. J Neurochemistry 1999. 73:2316-2320.
- Zambrzycka A, Kacprzak M. Apo lipo-protein E4 and A(beta)1-42 inhibit phosphoinositide biosynthesis in rat brain cortex. *Pol J Pharmacol* 2003. 55:911-913.
- Veinbergs I, Everson A, Sagara Y, Masliah E. Neurotoxic effects of apolipoprotein E4 are mediated via dysregulation of calcium homostatis. *J Neurosci Res* 2002. 67:379-387.
- Cohen AW, Combs TP, Scherer PE, Lisanti MP. Role of caveolin and caveolae in insulin signaling and diabetes. *Am J Physiol Endocrinol Metab* 2003. 285:E1151-E1160.
- Muller G, Frick W. Signaling via caveolin: involvement in the cross-talk between phosphoinositol glycans and insulin. *Cell Bi*ol Life Sci 1999. 56:945-970.
- Nystrom FH, Chen H, Cong LN, Li Y, Quon MJ. Caveolin-1 interacts with the insulin receptor and can differentially modulate insulin signaling in transfected Cos-7 cells and rat adipose cells. *Mol Endocrinol* 1999. 13:2013-2024.
- Fielding CJ, Bist A, Fielding PE. Caveolin mRNA levels are up-regulated by free cholesterol and down-regulated by oxysterols in fibroblast mono layers. *Proc Natl Acad Sci USA* 1997. 94:3753-3758.
- Suzuki N, Iwatsubo T, Odaka A, Ishibashi Y, Kitada C, Ihara Y. High tissue content of soluable beta 1-40 is linked to cerebral amyloid angiopathy. *Am J Pathol* 1994. 145:452-460.
- 52. Kojro E, Gimpl G, Lammich S, Marz W, Fahrenholz. Low cholesterol stimulates the non-amyloidogenic pathway by its effect on alpha-secretase ADAM 10. *Proc Natl Acad Sci USA* 1001. 98:5815-5820.
- 53. Kawarabayashi T, Shoji M, Younkin LH, Wen-Lang L, Dickson DW, Murakami T, Matsubara E, Abe K, Ashe KH, Younkin SG. Dimeric amyloid beta protein rapidly accumulates in lipid rafts followed by apolipoprotein E and phosphorylated tau accumulation in the Tg 2576 mouse model of Alzheimer's disease. *J Neurosci* 2004. 24:3801-3809.
- Lee S, Liyanage U, Bickel PE, Xia W, Lansbury PT Jr, Kosik KS. A detergent-insoluable membrane compartment contains A(beta) in vivo. *Nature Med* 1998. 4:730-734.
- 55. Lee, EB, Skovronsky DM, Abtahian F, Doms RW, Lee VM. Secretion and intracellular generation of truncated A(beta) in beta site amyloid-beta precursor protein-cleaving enzyme expressing human neurons. *J Biol Chem* 2003. 278:4458-4466.
- Solano DC, Sironi M, Bonfini C, Solerte SB, Govoni S, Racchi M. Insulin regulates soluble amyloid precursor protein release via phosphatidyl-inositol 3-kinase dependent pathway. *FASEB J* 2000. 14:1015-1022.
- Galvan V, Chen S, Lu D, Logvinova A, Goldsmith P, Koo EH, Bredesen DE. Caspase cleavage of members of the amyloid precursor family of proteins. *J Neurochem* 2002. 82:283-294.
- Li ZG, Zhang W, Sima AA. C-peptide enhances insulinmediated cell growth and protection against high glucose induced apoptosis in SH-SY5Y cells. *Diab Metab Res Rev* 2003. 19:375-385.
- Russel JW, Sullivan KA, Windebank AJ, Herrmann DN, Feldman EL. Neurons undergo apoptosis in animal and cell culture models of diabetes. *Neurobiol Dis* 1999. 6:347-363.

- Galvan V, Chen S, Lu D, Logvinova A, Goldsmith P, Koo EH, Bredesen DE. Caspase cleavage of members of the amyloid precursor family of proteins. J Neurochem 2003. 82:283-294.
- Lu DC, Rabizadek S, Chandra S, Shayya RF, Ellerby LM, Ye X, Salvesen GS, Koo EH, Bredesen DE. A second cyctotoxic proteolytic peptide derived from amyloid beta-protein percursor. *Nature Med* 2000. 6:397-404.
- 62. Tamagno E, Bardini P, Obbili A, Vitali A, Borghi R, Zaccheo D, Pronzato MA, Donni O, Smith MA, Perry G, Tabaton M. Oxidative stress increases expression of BACE in NT2 neurons. *Neurobiol Dis* 2002. 10:279-288.
- Bajt MC, Ho YS, Vonderfecht SL, Jaeschke H. Reactive oxygen as modulator of TNF and fas receptor mediated apoptosis in vivo: studies with glutathione peroxidase-deficient mice. *Antioxid Redox Signal* 2002. 4:733-740.
- Kamiya H, Zhang W, Sima AA. Apoptotic stress is counterbalanced by survival elements preventing programmed cell death of DRG's in subacute type 1 diabetic BB/Wor-rats. *Diabetes* 2003. 54:3288-3295.
- 65. Cheng C, Zochodne DW. Sensory neurons with activated caspase-3 survive long-term experimental diabetes. *Diabetes* 2003. 52:2363-2371.
- Johnson GV, Bailey CD. The p38 MAP kinase signaling pathway in Alzheimer's disease. *Exp Neurol* 2003. 183:262-268.
- Petanceska SS, Gandy S. The phosphotidylinositol 3-kinase inhibitor Wortmannin alters the metabolism of the Alzheimer's amyloid precursor protein. J Neurochem 1999. 73:2316-2320.
- Solano DC, Sironi M, Bonfini C, Solarte SB, Govani S, Racchi M. Insulin regulates soluble amyloid precursor protein release via phosphatidyl-inositol 3-kinase dependent pathway. *FASEB J* 2000. 14:1015-1022.
- 69. Xu H, Sweeney D, Greengard P, Gandy S. Metabolism of Alzheimer beta-amyloid precursor protein: regulation by protein kinase A in intact cells and in a cell-free system. *Proc Natl Acad Sci USA* 1996. 93:4081-4084.
- 70. Puig B, Gomez-Isla T, Ribe E, Candrado M, Torrejon-Escribano B, Dalfo E, Ferrer I. Expression of stressactivated kinases c-JunN-terminal kinase (SAPK/JNK-P) and p38 kinase (p38-P), and tau hyperphosphorylation in neurites surrounding betaA plaques in APPTg2576 mice. *Neuropath Appl Neurobiol* 2004. 30:491-502.
- Selkoe DJ. Alzheimer's disease: genes, proteins and therapy. *Physiol Rev* 2001. 81:741-766.
- Nixon RA, Wegiel J, Kumar A, Yu WH, Peterhoff C, Cataldo A, Cuervo AM. Extensive involvement of autophagy in Alzheimer disease: an immuno-electron microscopy study. J Neuropath Exp Neurol 2005. 64:113-122.
- Cotman CW, Poon WW, Rissman RA, Blurton-Jones M. The role of caspase cleavage of tau in Alzheimer disease neuropathology. J Neuropath Exp Neurol 2005. 64:104-112.
- 74. Lee VM, Goedert M, Trojanowski JQ. Neurodegenerative tauopathies. *Annu Rev Neurosci* 2001. 24:1121-1159.
- 75. Grundke-Iqbal I, Iqbal K, Tung YC, Quinlan M, Wiesniewski HM, Binder LI. Abnormal phosphorylation of the microtubule-associated protein tau (tau) in Alzheimer cytoskeletal pathology. *Proc Natl Acad Sci USA* 1986. 83:4913-4917.
- 76. Gamblin TC, Chen F, Zambrano A, Abraha A, Lagalwar S, Guillozet AL, Lu M, Fu Y, Garcia-Sierra F, La Pointe N, Miller R, Berry RW, Binder LI. Caspase cleavage of tau: linking amyloid to neurofibrillary tangles in Alzheimer's disease. *Proc Natl Acad Sci USA* 2003. 100:10032-10037.

- Fasulo L, Ugolini G, Visintin M. The neuronal microtubule associated protein tau is a substrate for caspase-3 and an effector of apoptosis. *J Neurochem* 2000. 75:624-633.
- Guo H, Albrecht S, Bourdeau M, Petzke T, Bergeron C, LeBlanc AC. Active caspase 6 and caspase-6-cleaved tau in neuropil threads, neuritic plaques and neurofibillary tangles of Alzheimer's disease. *Am J Pathol* 2004. 165:523-531.
- Rohn TT, Head E, Su JH. Correlation between caspase activation and neurofibrillary tangle formation in Alzheimer's disease. *Am J Pathol* 2001. 158:189-198.
- Su JH, Zhao M, Anderson AJ. Activated caspase-3 expression in Alzheimer's and aged control brains. Correlations with Alzheimer pathology. *Brain Res* 2001. 898:350-357.
- Lian Q, Ladner CJ, Magnuson D, Lee JM. Selective changes of calcineurin (protein phosphotase 2B) activity in Alzheimer's disease cerebral cortex. *Exp Neurol* 2001. 167:158-165.
- 82. Horndane M, Sambo AV, Delabel P, Begard S, Violleau A, Delacourte A, Bertrand P, Benavides J, Bree L. Mitotic-

like tau phosphorylation by p25-Cdk5 kinase complex. J Biol Chem 2003. 278:34026-34034.

- Phiel CJ, Wilson CA, Lee VM, Klein PS. GSK-3beta regulates production of Alzheimer's disease amyloid-beta peptides. *Nature* 2003. 423:435-439.
- Hong MF, Lee VM. Insulin and insulin-like growth factor-1 regulate tau phosphorylation in cultured human neurons. *J Biol Chem* 1997. 272:19547-19553.
- Cross DAE, Alessi DR, Cohen P, Andjelkovich M, Hemmings BA. Inhibition of glycogen synthase kinase-3 by insulin mediated protein kinase. *Nature* 1995. 378:785-789.
- 86. Hoyer S. Glucose metabolism and insulin signal transduction in Alzheimer's disease. *Eur J Pharmacol* 2004. 490:115-125.
- 87. Mandelkow EM, Drewes G, Biernet J, Gustke N, Van Lint J, Vandenheede JR, Mandekow E. Glycogen synthase-3 and the Alzheimer-like state of microtubule-associated protein tau. *FEBS Lett* 1992. 314:215-221.