

Autoimmune Diabetes: Ongoing Development of Immunological Intervention Strategies Targeted Directly Against Autoreactive T Cells

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Abstract

It is well known that autoimmunity associated with the onset of insulin-dependent diabetes mellitus (IDDM) involves the generation of autoreactive T and B cells. The findings that diabetics mount humoral and cellular immune responses against islet cell antigens (ICAs) have led to the testing of ICAs and their analogs as candidates for therapeutic agents for better treatment of IDDM at its prediabetic and diabetic stages. Apart from this type of approach, various immunological intervention strategies aimed at direct targeting of the autoreactive T cells have also been investi-

Introduction

DDM is a form of diabetes mellitus (DM) characterized by the dysfunction of the pancreas to produce insufficient or no insulin. This disorder is caused by autoimmune-mediated destruction of the pancreatic β -cells. Over the years, it has been observed that autoimmunity associated with IDDM involves the participation of both B and T autoreactive lymphocytes. The evidence for B cell-mediated autoimmunity is based on the well reported findings that up to 98% of IDDM patients have antibodies against one or more of their own β -cell antigens. These include: Insulin [1]; the major of the 2 isoforms of glutamic acid decarboxylase (GAD) 65 [2]; two of the protein tyrosine phosphatases, insulinoma antigen-2 and insulinoma antigen-2 β (IA-2 and IA-2 β) [3, 4]; the heterogeneous islet cell cytoplasmic antigens (ICAs) [5, 6]. A minority of IDDM patients also have serum antibodies to a gated. The present review covers the ongoing aspects of these developments focusing on the preclinical findings made in nonobese diabetic (NOD) mice which have been commonly used as a disease model for human autoimmune diabetes. Other types of approaches involving the mobilization of regulatory T cells to indirectly control or modulate the pathological activity of autoreactive T cells will not be discussed within this scope.

Keywords: type 1 diabetes • autoimmunity • T cells • regulation • co-stimulation

glycosylated islet cell membrane antigen, GLIMA [7]. Recently, autoantibodies to other new antigens of protein tyrosine phosphatases, IA-2/ICA512 and IA-2 β /phogrin, expressed by pancreatic islet cells, have been detected [8].

The generation of autoantibodies to islet cells can be observed for as many as 10 years prior to the onset of clinical diabetes [9]. Despite this observation, the existence of autoantibodies is not solely sufficient to cause IDDM development. This conclusion is based on the finding that infants born of antibody positive IDDM mothers can remain free of disease despite the existence of serum autoantibodies to insulin, GAD and other islet cell antigens [10]. On the other hand, persons with severe genetic B cell deficiency can still develop IDDM [11]. Generally, the level of autoantibodies correlates with the state of β -cell destruction [12, 13]. As such, they can therefore serve as indicators for autoimmune diabetes. A low level of GAD-specific autoantibodies is associated with a slow breakdown of β -cell function, while a high level of autoantibodies to IA-2 together with the maturation of autoantibody responses elicited against ICAs or GAD are signs for more severe and imminent β -cell failure [14].

Today it is well-known that IDDM is mediated by autoreactive T cells. The most direct indication of this finding is due to the examination of biopsy tissues obtained near the time of IDDM diagnosis, which show that the islets are infiltrated with activated T cells, primarily of the CD8+ population but also, to a lesser extent, CD4+ cells and macrophages as well [15-18]. The association of IDDM with the major histocompatibility complex (MHC)-associated susceptibility gene locus, IDDM1, is also well reported [19]. Recurrence of organ-specific autoimmunity is responsible for β -cell destruction in diabetics transplanted with a pancreatic graft donated by their discordant, nondiabetic monozygotic twins [20]. Furthermore, IDDM is transferable to non-diabetics given bone marrow transplant donated by diabetic HLA-identical siblings, or allogeneic donors [21, 22].

Autoreactive CD4+ cells of the Th1 subset are potentially capable of directly and indirectly causing islet damage: Directly via the release of cytotoxic mediators such as nitric oxide or oxygen radicals [23], and indirectly through the secretion of IL-2 and IFN- γ by activating autoreactive CD8+ T cells and macrophages leading to their infiltration of the islets [24]. In this regard, characterization and quantitation of autoreactive T cells in humans are important for the development of an improved diagnosis of IDDM, and intervention strategies for arresting disease progression. However, direct detection of autoreactive T cells in IDDM is more difficult than the detection of autoantibodies. The reason is that CD4+ and CD8+ autoreactive T cells generated in the course of IDDM development are only present at very low frequencies in the circulation of subjects with recent disease onset [25, 26]. Assays dependent on in vitro expansion to allow the detection of autoreactive CD4+ T cells in the pool of peripheral blood leucocytes (PBL) of diabetics are used in most studies. When employing in vitro proliferation assays, PBL of individuals with recent onset of IDDM respond to human insulin [27], a spectrum of islet cell antigens [28-30], and GAD [31]. Regarding detection, GAD-specific autoreactive T cells can be generated and cloned from peripheral T cells of recent onset IDDM patients who are carrying the diseasesusceptible HLA-DR alleles [32]. Furthermore, endogenous GAD fragments presented by IDDM- associated HLA class II molecules can be isolated [33]. The identification of HLA-DR-restricted GAD epitopes facilitates the *in vitro* expansion of peripheral GAD-specific T cells from individuals with IDDM atrisk and subjects with IDDM and the subsequent demonstration of the $\alpha\beta$ TCR specificity of the autoreactive T cells against the corresponding ligand using the HLA-class II tetramer analysis procedure [34].

Some laboratories focus on the characterization of diabetogenic CD8+ T cells. So far, autoreactive CD8+ T cells have been detected in humans against two β cell antigens, namely GAD 65 and preproIAPP (precursor human islet amyloid polypeptide protein), which are cosecreted with insulin in subjects recently diagnosed with IDDM. GAD 65-specific cytotoxic T cells (CTLs) carrying the disease-associated allele, HLA-A2, following in vitro expansion with a HLA-A2 binding peptide, have been generated from PBL of these individuals [35]. A recent study describes the presence of an autoreactive CD8+ subset in the circulation of recently diagnosed patients that recognize a 9 amino acid long immunodominant epitope of preproIAPP in the context of HLA-A2 using an IFN-ybased ELISPOT assay [36]. The feasibility to detect and quantitate circulating autoreactive T cells directly at early disease onset can undoubtedly serve as a valuable tool for improved diagnosis of IDDM, and the development of better tolerogens which can be used to arrest IDDM onset.

Immunological intervention strategies

Normally, IDDM is diagnosed by the time 70-90% of the patient's islet cells are already destroyed. Over the years, immunological intervention strategies have been tested for their effectiveness at the prediabetic stage of the disease to prevent its subsequent onset, or at the early stage of the disease to halt further assault on the pancreatic islets in order to allow the remaining function of the organ to be preserved. A common approach to these strategies is to target the autoreactive T cells directly. Several of these developments do work primarily in the NOD mouse model of human diabetes, some have reached the clinical trial stage. The remaining part of this review will discuss the ongoing development of these strategies and the rationale behind why they are aimed at interfering with the activity and function of autoreactive T cells believed to be involved in causing IDDM.

Intervention at prediabetic stage

1. Autoantigen (insulin and GAD)-induced tolerance

In view of the detection of autoreactive T cells showing specificities against islet cell antigens, intervention strategies involving the administration of soluble and formulated islet cell antigens aim at tolerizing these autoreactive T cells. For this type of approach, three antigens, namely insulin, the glutamic decarboxylase (GAD65) and heat shock protein (hsp) 60, have attracted the most attention since diabetics are known to mount immune responses against them. It has been shown in preclinical studies that administration of the native form or analogs of these antigens induce T cell mediated regulatory mechanisms capable of preventing the development of autoimmune diabetes in the NOD mice. Thus, prediabetic animals treated with insulin are less susceptible to subsequent IDDM development [37, 38]. Induction of insulin-specific regulatory T lymphocytes is protective since untreated mice that accept T cells from those administered with insulin are also protected from disease development. The protection mechanism can be attributed to the production of immuno-suppressive cytokines, such as IL-4, IL-10 and TGF- β that exert negative regulation of the diabetogenic Th1 cells [39].

Prediabetic NOD mice subcutaneously injected with a recombinant version of GAD65 show a cellular response dominated by the secretion of IFN- γ by the GAD65-specific Th1 cells. Interestingly, additional injection of GAD65 is able to change the response deviating towards the production of Th2 cytokines, IL-4 and IL-10 that suppress the pathogenic role mediated by the autoreactive Th1 T cells [40].

In 1998 a previous pilot study reported that insulin therapy is effective in preventing IDDM onset in a significant number of high-risk individuals [41]. However, clinical trials conducted as part of the Diabetes Prevention Trial-Type 1 (DPT-1) have recently reported that both low-dose insulin administration using a similar protocol to that in the pilot study, as well as the oral intake of insulin capsules are unable to protect high-risk subjects from developing the disease [42]. The reason for this outcome can only be postulated at the present time to be attributable to several factors, such as a difference in the study groups or insufficient administration of insulin for making a difference [43]. It has not been shown in a study conducted by the American Diabetes Association that intervention with the GAD65 molecule (Diamyd) is able to offer clear clinical benefit. The immunogen is currently being evaluated for its effectiveness to stop the onset of LADA (latent autoimmune diabetes of adults) [44].

2. Tolerance induction with second generation autoantigens (insulin & GAD peptides)

The function of a subset of pathogenic T cells can be modulated via the engagement of their $\alpha\beta$ TCR with a natural (depending on the mode of administration) or an artificial ligand in association with the respective MHC class I or II molecule. A fragment comprising the amino acid residues 9-23 of the human insulin Bchain contains a promiscuous immunodominant T cell epitope that can be recognized by CD4+ autoreactive T cells of diabetics and the NOD mice. A proportion of CD4+ T cells infiltrating the diseased islets of the diabetic mice responds to peptide B:9-23, suggesting that autoimmunity directed against this site of the human insulin B-chain significantly contributes to IDDM [45, 46]. Subcutaneous injection of pepitde B:9-23 formulated in IFA or intranasal administration of the peptide is capable of conferring protection against IDDM development in prediabetic NOD mice. The protection is associated with modulation of autoreactive T cell function as judged by their reduced capability to proliferate to peptide B:9-23 [47, 48].

More recently, an artificial version of peptide B:9-23, NBI-6024, containing alanine substitutions at positions 16 and 19 of the parent fragment has been tested for its effectiveness in arresting IDDM. The altered ligand inhibits the proliferation of B:9-23-specific autoreactive T cells obtained from the NOD mice. Furthermore, subcutaneous injection of soluble NBI-6024 is capable of protecting both prediabetic and diabetic NOD mice from disease development. The mechanism of protection has been attributed to immune-deviation based on the observation that NBI-6024 preferentially induces T cells of the vaccinated animals to produce Th2 immunosuppressive cytokines, IL-4 and IL-10 (49).

In tests NBI-6024 shows sufficient safety levels to reach a phase I clinical trial. It will be interesting to know whether the strategy adopted to target an immunodominant population of diabetogenic T cells may prove to be more effective in preventing IDDM development than its native parent protein. In this regard, a separate report has recommended that the soluble immunogen would be best administered at low doses to avoid/minimize the development of anaphylaxis [50].

In addition to its ability to modulate the function of autoreactive CD4⁺ T cells, the fragment comprising

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the amino acid residue 15-23 within NBI-6024 also appears to contain an immunodominant murine MHC class I-restricted T cell epitope. Murine islet resident diabetogenic CD8⁺ T cells are able to recognize peptide B:15-23 [51]. Despite the fact that an efficacy study has not been performed with peptide B:15-23, the results of a recent study suggest that vaccination with an altered influenza virus haemagglutinin (HA) MHC class I-restricted ligand can result in modulating the activity of diabetogenic HA-specific CD8⁺ T cells in the prevention of diabetes onset in a transgenic mouse model. Efficacy of the therapy is associated with decreased infiltration of the pathogenic T cells to the islets, and reduced cytotoxic potential of the effector cells [52].

GAD65 epitopes have similarly been investigated for their capabilities to arrest IDDM in the NOD mouse. The rationale for this approach is based on the finding that spontaneous disease onset in the diabetic animal is associated with T cell reactivities directed against various epitopes of GAD65. Intervention of GAD65-specific autoreactivity at the prediabetic stage may therefore be an effective means for IDDM prevention. We already know that certain epitopes of GAD65 are capable of arresting IDDM in NOD mice. GAD65 246-266, but not GAD65 509-528 or GAD65 524-543 peptide, given in PBS to pregnant animals using the ip (intraperitoneal) route induce tolerance of the offspring in terms of resistance against diabetes development. Surprisingly, vaccination with a mixture of GAD65 509-528 and GAD65 524-543 peptides, but not a triple combination of GAD65 509-528, GAD65 524-543 and GAD65 246-266 is also effective. The reason for this outcome is unclear at the moment. Overall, more investigation is needed to establish a clearer picture of the effectiveness of GAD peptides in IDDM prevention [53-55].

3. Antibody-induced modulation

Another approach to prevent neonatal NOD mice from developing IDDM involves the administration of a monoclonal antibody directed against the CD3 complex non-covalently associated with the $\alpha\beta$ TCR expressed on conventional CD4 and CD8 subsets of T cells. Upon engagement of the $\alpha\beta$ TCR with its foreign ligand, or ligand derived from self antigen (in the case of autoreactive T cells), CD3 can mediate signal transduction events leading to the activation of T cells. Thus, binding of an anti-CD3 antibody to CD3 can be expected to affect the activity of autoreactive T cells in causing damage to the pancreatic β -cells. Preclinical

studies demonstrated that young streptozocin (STZ)induced diabetic CD1 mice administered with the nonmitogenic [F(ab')2] fragment of a monoclonal antibody directed against the ε chain of human CD3 are prevented from developing insulitis. Protection is attributed partly to the depletion of some T cells, and partly to the antibody being capable of inducing anergy to a significant proportion of T cells as judged by their failure to produce the Th1 type of cytokines such as IL-2 and IFN- γ [56].

4. Antibody-blocking of autoreactive T cell activation

Similar to conventional T lymphocytes, full activation of autoreactive CD4+ and CD8+ T cells requires both the engagement of its $\alpha\beta$ TCR with an antigenic peptide in association with the respective MHC class I or II molecules, as well as a second signal provided by co-stimulation. A co-stimulatory signal will be generated by the interaction of CD28 molecules expressed on naïve T cells with its ligands, B7-1 (CD80) and B7-2 (CD86), presented on APCs. Of the two ligands, B7-2 plays a more dominant role in interacting with CD28 to trigger T cell activation [57, 58]. In NOD mice, the engagement of CD28 with B7-1 and B7-2 are critical for the control of spontaneous diabetes development. Animals without CD28, and B7-1 with B7-2, have a reduced number of CD4+CD25+ regulatory T cells and succumb to clinical disease faster than their wild-type counterpart [59].

Reagents that bind B7-1 and/or B7-2 have been evaluated for their effectiveness to block the activation of autoreactive T cells in the course of IDDM prevention in NOD mice. Indeed, prediabetic NOD mice of 2-3 weeks of age given multiple doses of an anti-B7-2 monoclonal antibody over a period of 8 weeks are protected from subsequent development of diabetes. Animals subjected to a similar regimen by injecting them with a fusion protein, CTLA-4Ig comprising the extracellular domain of CTLA-4 and the fragment of human IgG1, which binds to B7-1 and B7-2 to inhibit costimulation, are also protected from the onset of IDDM [60-62]. Fine analysis of the NOD mice subjected to the entire course of anti-B7-2 and CTLA-4Ig treatments identifies that the reagents inhibit the development of clinical disease rather than insulitis because animals not treated on weeks 6 to 8 (final phase of therapy) succumb to IDDM [63]. Surprisingly, the administration of an anti-B7-1 monoclonal antibody mediates an opposite effect by triggering an accelerated onset of the disease not only in the diabetes-prone female mice, but in the normally diabetes-resistant

male mice as well. Apparently, disease exacerbation is due to the presence of pancreatic T cells displaying higher expression of CD69 indicating that T cells are in a higher activated state than those found in the pancreas of animals treated with the anti-B7-2 antibody and CTLA-4Ig. The contrasting outcome obtained from these studies can be attributed to the differential expression of B7-2 and B7-1 on lymphoid cells. Given that B7-2 is expressed more rapidly than B7-1 on APCs when they become activated [64], blocking of B7-2 seems essential for inhibiting its interaction with CD28 to arrest diabetes development. Results obtained from studies of B7-2 knock-out NOD mice similarly show that co-stimulation in the total absence of B7-2 participation is capable of delaying insulitis development until late in the adulthood of the animals. This is associated with a redistribution of pathogenic T cells to the spleens instead of the islets of the mice. Despite the observed clinical benefit, T cell differentiation and activation without B7-2 in the B7-2 knock outs generates neuropathy characterized by severe inflammation of the peripheral nerves which precedes insulitis development [64]. On the other hand, blocking of B7-1 expressed on activated T cells would inhibit its interaction with CTLA-4 leading to a failure in generating a regulatory signal to control IDDM development.

Overall, co-stimulatory pathways leading to the regulation of T cell activation and function are complex, and further in-depth investigation is required to modify them for the intervention of IDDM.

Intervention at the stage of overt diabetes

1. Autoantigen (hsp)-induced tolerance

In the course of spontaneous diabetes development, NOD mice develop an immunodominant autoreactive T cell response directed against a fragment comprising the amino acid residues 437-460 of hsp 60. T cells generated against a peptide, p277, encoding this sequence of hsp60 are diabetogenic based on the observation that recipient mice adoptively transferred with the cells turn diabetic [65]. Thus, targeting this p277-specific subset of T cells to modulate its pathological function may prove useful for arresting IDDM. The spontaneous development of diabetes in adult NOD mice is preventable by a single injection of a hsp60 peptide, p277, formulated in IFA (Incomplete Freund's Adjuvant). Efficacy is associated with a reduced infiltration of IFN-y-secreting T cells to the islets of the p277 treated mice [66, 67]. Follow up studies have demonstrated that T cells of p277 injected NOD mice secrete disproportionate amounts of IL-4 and IL-10 to down-regulate Th1 responses elicited not only to the immunizing antigen, but to GAD65 and insulin as well. Apart from the p277 encoded epitope, another study reports the discovery of another hsp60 epitope represented by the sequence 166-185 of the protein that is effective in arresting diabetes in the NOD mice [68].

Peptide p277 is now being tested in a double blind phase II clinical trial. Subjects with recent onset of IDDM subcutaneously injected with two doses of the peptide maintain their insulin secretion based on Cpeptide measurement, and reduce their need for exogenous insulin as compared to the placebo group [69].

2. Antibody-induced modulation

Several antibodies directed against lymphocytic antigens have been assessed for their effectiveness in halting disease progression in NOD mice. Apart from its effectiveness in preventing IDDM onset, multiple administration of the human CD3e chain-specific monoclonal antibody is capable of conferring longterm protection in adult NOD mice with overt diabetes. Adult NOD mice injected daily with a low-dose regimen of 5.0 µg of the antibody for 5 days are protected from accelerated IDDM development induced by cyclophosphamide. The same therapeutic application is also effective in arresting spontaneous diabetes in adult animals for a significantly long period of time. Apparently, protection is not associated with the clearance of diabetogenic CD8+ cells in the pancreatic islets of the animals [70]. The efficacy generated by anti-CD3 treatment is mediated primarily by TGF- β that is produced by a subset of CD4+CD25- regulatory T cells, as shown in a follow-up study using a similar protocol with a high-dose regimen of 50.0 µg of the antibody [71]. Protection provided by anti-CD3 antibody administration is also attributed to the induction of anergy in autoreactive CD8⁺ cells [72]. The safety of a humanized version of the murine anti-CD3 monoclonal antibody, hOKT3y1Ala-Ala, has been evaluated and subsequently approved for phase one trials in patients with recent-onset of IDDM [73]. So far, the results obtained from this trial are encouraging as judged by the majority of the treated patients being able to maintain and improve their insulin secretion. Interestingly, one parameter associated with the observed efficacy is an increase in the number of CD8+

cells in the treated subjects [74]. The regulatory/effector function of these cells remains to be investigated.

NOD mice can also be protected from spontaneous diabetes development when they are treated with the non-depleting anti-CD4 monoclonal antibody, YTS177.9 [75, 76]. The protection mechanism seems to be due to the antibody acting directly against the autoreactive T cells since mice with infiltrates of CD4+ diabetogenic T cells in their islets are also protected upon the administration of the antibody. Follow-up studies suggest that YTS177.9 binding affects autoreactive T cells by altering their capability to secrete proinflammatory cytokines [76, 77].

3. Antibody blockade of autoreactive T cell migration

Antibodies capable of interfering with the trafficking of autoreactive mononuclear cells to the pancreatic islets have been evaluated for arresting the ongoing diabetes in the NOD mice. Migration of lymphocytes, including autoreactive T cells, requires chemoattractants and their adhesion to endothelial cells for migration from the circulation into the tissues and organs [78]. The integrin, $\alpha 4\beta 7$, is expressed on the infiltrating mononuclear cells during the process of insulitis development [79]. A monoclonal antibody against the $\alpha 4$ subunit of the heterodimer, $\alpha 4\beta 7$, or VCAM-1 (the ligand for the integrin), is able to block the entry of insulitis-causing mononuclear cells to the islet of the diabetic mice [80, 81].

CD43 (leucosialin), expressed as a 95-135 kDa highly glycosylated protein on T cells, is an adhesion molecule [82]. The administration of a monoclonal antibody against CD43 is effective in preventing the onset of diabetes in newborn and adult NOD mice [83]. Protection is attributed to the inhibition of autoreactive T cells migrating to the pancreas.

4. Antibody-induced deletion

Results of earlier studies show that the administration of mono-specific antibodies against MHC class I and CD8 molecules are effective in preventing further β -cell destruction in the IDDM-prone male adult

References

- Atkinson MA, Maclaren NK, Riley WJ, Winter WE, Fisk DD, Spillar RP. Are insulin autoantibodies markers for insulin-dependent diabetes mellitus? *Diabetes* 1986. 35:894-898.
- Atkinson MA, Kaufman DL, Newman D, Tobin AJ, Maclaren NK. Islet cell cytoplasmic autoantibody reactivity to glutamate decarboxylase in insulin-dependent diabetes. J Clin

NOD/Shi/Kbe and female NOD mice, respectively [84, 85]. These observations are associated with severe T cell depletion as judged by the absence of CD8⁺ cells in the spleen and islets of the experimental mice. There has been no follow-up in the further development of these reagents for the prevention of IDDM in humans.

5. $\alpha\beta$ TCR-targeted tolerance induction

A separate approach using a fusion protein comprising a MHC class II molecule linked to a specific altered ligand has been tested preclinically for its effectiveness in preventing IDDM progression. The ligand of a chimeric construct, BDC2.5-IAg7, binds well to a subset of CD4⁺ diabetogenic T cells (BDC2.5). Administration of BDC2.5-IAg7 is able to confer protection to NOD.BDC2.5 TCR transgenic mice transferred with the BDC2.5 cells. Efficacy is attributed to the ability of the fusion protein to induce death and anergy to some of the BDC2.5 cells and to modulate the remaining ones to secrete more of the Th2 type of anti-inflammatory cytokines [86].

Summary

The advent of our understanding of IDDM has opened up a variety of opportunities for us to utilize in the search for better treatment and prevention of the disease. Improved detection of autoreactive T cells in patients with IDDM may offer better and earlier diagnosis of the disease. Improved characterization of diabetogenic T cells can be applied to the development of more effective intervention strategies to arrest the spontaneous progression of IDDM. Despite the failure of the insulin trials, results revealed by the clinical trial conducted with the human anti-CD3 specific monoclonal antibody are encouraging, and there are promising findings from other preclinical studies as well. Although not covered in this review, it needs to be mentioned here that the role of regulatory T cells in the control of IDDM is also becoming clearer, and ways to stimulate them to exert positive immuno-regulation for protection against IDDM are emerging. Let us hope that some more of these strategies will make their way through to clinical assessments.

Invest 1993. 91:350-356.

Lu J, Li Q, Xie H, Chen ZJ, Borovitskaya AE, Maclaren NK, Notkins AL, Lan MS. Identification of a second transmembrane protein tyrosine phosphatase, IA-2beta, as an autoantigen in insulin-dependent diabetes mellitus: precursor of the 37-kDa tryptic fragment. *Proc Natl Acad Sci U S A* 1996. 93:2307-2311.

^{4.} Lan MS, Wasserfall C, Maclaren NK, Notkins AL. IA-2, a

transmembrane protein of the protein tyrosine phosphatase family, is a major autoantigen in insulin-dependent diabetes mellitus. *Proc Natl Acad Sci U S A* 1996. 93: 6367-6370.

- Gorus FK, Goubert P, Semakula C, Vandewalle CL, De Schepper J, Scheen A, Christie MR, Pipeleers DG. IA-2autoantibodies complement GAD65-autoantibodies in newonset IDDM patients and help predict impending diabetes in their siblings. *Diabetologia* 1997. 40:95-99.
- Strebelow M, Schlosser M, Ziegler B, Rjasanowski I, Ziegler M. Karlsburg type I diabetes risk study of a general population: frequencies and interactions of the four major type I diabetes associated autoantibodies studied in 9419 schoolchildren. *Diabetologia* 1999. 42:661-670.
- Aanstoot HJ, Kang SM, Kim J, Lindsay LA, Roll U, Knip M, Atkinson M, Mose-Larsen P, Fey S, Ludvigsson J, et al. Identification and characterization of glima 38, a glycosylated islet cell membrane antigen, which together with GAD65 and IA2 marks the early phases of autoimmune response in type 1 diabetes. *J Clin Invest* 1996. 97:2772-2783.
- Kawasaki E, Yu L, Rewers MJ, Hutton JC, Eisenbarth GS. Definition of multiple ICA512/phogrin autoantibody epitopes and detection of intramolecular epitope spreading in relatives of patients with type 1 diabetes. *Diabetes* 1998. 47:733-742.
- Luhder F, Schlosser M, Mauch L, Haubruck H, Rjasanowski I, Michaelis D, Kohnert KD, Ziegler M. Autoantibodies against GAD65 rather than GAD67 precede the onset of type 1 diabetes. *Autoimmunity* 1994. 19:71-80.
- Yu L, Eisenbarth GS. Type 1 diabetes: Molecular, cellular and clinical immunology. www.uchsc.edu/misc/diabetes/ oxch10.html - 67k, last updated Feb 2004.
- Martin S, Wolf-Eichbaum D, Duinkerken G, Scherbaum WA, Kolb H, Noordzij JG, Roep BO. Development of type 1 diabetes despite severe hereditary B-lymphocyte deficiency. N Engl J Med 2001. 345:1036-1040.
- Irvine WJ, McCallum CJ, Gray RS, Campbell CJ, Duncan LJ, Farquhar JW, Vaughan H, Morris PJ. Pancreatic isletcell antibodies in diabetes mellitus correlated with the duration and type of diabetes, coexistent autoimmune disease, and HLA type. *Diabetes* 1997. 26:138-147.
- Riley WJ, Maclaren NK, Krischer J, Spillar RP, Silverstein JH, Schatz DA, Schwartz S, Malone J, Shah S, Vadheim C, et al. A prospective study of the development of diabetes in relatives of patients with insulin-dependent diabetes. N Engl J Med 1990. 323:1167-1172.
- Borg H, Gottsater A, Landin-Olsson M, Fernlund P, Sundkvist G. High levels of antigen-specific islet antibodies predict future beta-cell failure in patients with onset of diabetes in adult age. J Clin Endocrinol Metab 2001. 86:3032-3038.
- 15. Bottazzo GF, Dean BM, McNally JM, Mackay EH, Swift PGF, Gamble DR. In situ characterization of autoimmune phenomena and expression of HLA molecules in the pancreas in diabetes insulitis. *N Engl J Medicine* 1985. 313:353-360.
- Hanninen A, Jalkanen S, Salmi M, Toikkanen S, Nikolakaros G, Simell O. Macrophages, T cell receptor usage, and endothelial cell activation in the pancreas at the onset of insulin-dependent diabetes mellitus. *J Clin Invest* 1992. 90:1901-1910.
- 17. Itoh N, Hanafusa T, Miyasaki A, Miyagawa J, Yamura S, Inada M, Kawata S, Tarui S, Kono N, Matsuzawa Y. Mononuclear cell infiltration and its relation to the expression of major histocompatibility complex antigens and adhesion molecules in pancreas biopsy specimens from newly diagnosed insulin-dependent diabetes mellitus patients. J Clin Invest 1993.

92:2313-2322.

- Imagawa A, Hanafusa T, Tamura S, Moriwaki M, Itoh N, Yamamoto K, Iwahashi H, Yamagata K, Waguri M, Nanmo T, et al. Pancreatic biopsy as a procedure for detecting in situ autoimmune phenomena in type 1 diabetes. *Diabetes* 2001. 50:1269-1273.
- Froguel P. Genetics of type 1 insulin-dependent diabetes mellitus. *Horm Res* 1997. 48(Suppl 4):55-57.
- Sutherland DE, Sibley R, Xu XZ, Michael A, Srikanta AM, Taub F, Najaran J, Goetz FC. Twin-to-twin pancreas transplantation: Reversal and reenactment of the pathogenesis of type 1 diabetes. *Trans Assoc Am Physicians* 1984. 97:80-87.
- Lampeter EF, Homberg M, Quabeck K, Schaefer UW, Wernet P, Bertrams J, Gorsse-Wilde H, Gries FA, Kolb H. Transfer of insulin-dependent diabetes between HLA-identical siblings by bone marrow transplantation. *Lancet* 1993. 341:1243-1244.
- Marmont AM. Stem cell transplantation for severe autoimmune disorders, with special reference to rheumatic diseases. J Rheumatol 1997. 48(Suppl):13-18.
- 23. Held W, MacDonald HR, Weissman IL, Hess MW, Mueller C. Genes encoding tumor necrosis factor alpha and Granzyme A are expressed during development of autoimmune diabetes. *Proc Natl Acad Sci. U S A* 1990. 87:2239-2243.
- Jean-Michel D, Burger D. Cytokines and direct cell contact in synovitis: Relevance to therapeutic intervention. *Arthritis Res* 1999. 1:17-20.
- 25. Tisch R, McDevitt H. Insulin-dependent diabetes mellitus. *Cell* 1996. 85:291-297.
- Notkins AL, Lernmark A. Autoimmune type 1 diabetes: Resolved and unresolved issues. J Clin Invest 2001. 108:1247-1252.
- Keller RJ. Cellular immunity to human insulin in individuals at high risk for the development of type 1 diabetes mellitus. J Autoimmunity 1994. 3:321-327.
- Roep BO, Kallan AA, Duinkerken G, Arden SD, Hutton JC, Bruining GJ, de Vries RR. T-cell reactivity to beta-cell membrane antigens associated with beta-cell destruction in IDDM. *Diabetes* 1995. 44:278-283.
- Brooks-Worrell BM, Starkebaum GA, Greenbaum C, Palmer JP. Peripheral blood mononuclear cells of insulindependent diabetes patients respond to multiple islet cell proteins. J Immunol 1996. 157:5668-5674.
- Mayer A, Rharbaoui F, Thivolet C, Orgiazzi J, Madec AM. The relationship between peripheral T cell reactivity to insulin, clinical remissions and cytokine production in type 1 (insulin-dependent) diabetes mellitus. J Clin Endocrinol Metab 1999. 84:2419-2424.
- Atkinson MA, Kaufman DL, Campbell L, Gibbs KA, Shah SC, Bu DF, Erlander MG, Tobin AJ, Maclaren NK. Response of peripheral-blood mononuclear cells to glutamate decarboxylase in insulin-dependent diabetes. *Lancet* 1992. 339(8791):458-459.
- 32. Endl J, Otto H, Jung H, Dreisbusch B, Donie F, Stahr P, Elbracht R, Schmitz G, Mein E, Hummel M, Ziegler AG, Wank R, Schendel DJ. Identification of Naturally Processed T Cell Epitopes from Glutamic Acid Decarboxylase Presented in the Context of HLA-DR Alleles by T Lymphocytes of Recent Onset IDDM Patients. J Clin Invest 1997. 99:2405-2415.
- 33. Nepom GT, Lippolis JD, White FM, Masewicz S, Marto JA, Herman A, Luckey CJ, Falk B, Shabanowitz J, Hunt DF, Engelhard VH, Nepom BS. Identification and modulation of a naturally processed T cell epitope from the diabetes-

associated autoantigen human glutamic acid decarboxylase 65 (hGAD65). *Proc Natl Acad Sci. U S A* 2001. 98:1763-1768.

- 34. Reijonen H, Novak EJ, Kochik S, Heninger A, Liu AW, Kwok WW, Nepom GT. Detection of GAD65-specific T-Cells by major histocompatibility complex class II tetramers in type 1 diabetic patients and at-risk subjects. *Diabetes* 2002. 51:1375-1382.
- 35. Panina-Bordignon P, Lang R, van Endert PM, Benazzi E, Felix AM, Pastore RM, Spinas GA, Sinigaglia F. Cytotoxic T cells specific for glutamic acid decarboxylase in autoimmune diabetes. J Exp Med 1995. 181:1923-1927.
- 36. Panagiotopoulos C, Qin H, Tan R, Verchere CB. Identification of a beta-cell-specific HLA class I restricted epitope in type 1 diabetes. *Diabetes* 2003. 52:2647-2651.
- Atkinson MA, Maclaren NK, Luchetta R. Insulitis and diabetes in NOD mice reduced by prophylactic insulin therapy. *Diabetes* 1990. 39:933-937.
- Zhang ZJ, Davidson L, Eisenbarth G, Weiner HL. Suppression of diabetes in nonobese diabetic mice by oral administration of porcine insulin. *Proc Natl Acad Sci. U S A* 1991. 88:10252-10256.
- 39. Hancock WW, Polanski M, Zhang J, Blogg N, Weiner HL. Suppression of insulitis in non-obese diabetic (NOD) mice by oral insulin administration is associated with selective expression of interleukin-4 and -10, tranforming growth factorbeta, and postaglandin-E. *Am J Path* 1995. 147:1193-1199.
- Plesner A, Worsaae A, Dyrberg T, Gotfredsen C, Michelsen BK, Pertersen JS. Immunization of diabetesprone or non-diabetes-prone mcie with GAD65 does not induce diabetes or islet cell pathology. J Autoimmunity 1998. 11:335-341.
- Fuchtenbusch M, Rabl W, Grassl B, Bachmann W, Standl E, Ziegler AG. Delay of type 1 diabetes in high risk, first degree relatives by parental antigen administration: The Schwabing Insulin Prophylaxis Pilot Trial. *Diabetologia* 1998. 41:536-541.
- 42. Marks JB. In pursuit of type 1 diabetes prevention. *Clinical Diabetes* 2002. 20:168-169.
- Masteller EL, Bluestone JA. Immunotherapy of insulindependent diabetes mellitus. *Curr Opin Immunol* 2002. 14:652-659.
- 44. Chase HP, Hayward AR, Eisenbarth GS. Clinical trials for the prevention of type I diabetes. www.uchsc.edu/misc/diabetes/oxch12.html
- Alleva DG, Crowe PD, Jin L, Kwok WW, Ling N, Gottschalk M, Conlon PJ, Gottlieb PA, Putnam AL, Gaur A. A disease-associated cellular immune response in type 1 diabetics to an immunodominant epitope of insulin. *J Clin Invest* 2001. 107:173–180.
- 46. Wegmann DR, Norbury-Glaser M, Daniel D. Insulinspecific T cells are a predominant component of islet infiltrates in pre-diabetic NOD mice. *Eur J Immunol* 1994. 24:1853-1857.
- Daniel D, Wegmann DR. Protection of nonobese diabetic mice from diabetes by intranasal or subcutaneous administration of insulin peptide B-(9-23). *Proc Natl Acad Sci U S A* 1996. 93:956-960.
- Daniel D, Wegmann DR. Intranasal administration of insulin peptide B:9-23 protects NOD mice from diabetes. *Proc Natl Acad Sci U S A* 1996. 93:956-960.
- 49. Alleva DG, Gaur A, Jin L, Wegmann D, Gottlieb PA, Pahuja A, Johnson EB, Motheral T, Putnam A, Crowe PD, et al. Immunological characterization and therapeutic activity of an altered-peptide ligand, NBI-6024, based on the

immunodominant type 1 diabetes autoantigen insulin B-chain (9-23) peptide. *Diabetes* 2002. 51:2126-2134.

- Liu E, Moriyama H, Abiru N, Miao D, Yu L, Taylor RM, Finkelman FD, Eisenbarth GS. Anti-peptide autoantibodies and fatal anaphylaxis in NOD mice in response to insulin self peptides B:9-23 and B-13-23. *J Clin Invest* 2002. 110:1021-1027.
- Wong FS, Karttunen J, Dumont C, Wen L, Visintin I, Pilip IM, Shastri N, Pamer EG, Janeway CA. Identification of an MHC class I-restricted autoantigen in type 1 diabetes by screening an organ-specific cDNA library. *Nat Med* 1999. 5:1026-1031.
- Hartemann-Heurtier A, Mars LT, Bercovici N, Desbois S, Cambouris C, Piaggio E, Zappulla J, Saoudi A, Liblau RS. An altered self-peptide with superagonist activity blocks a CD8 mediated mouse model of type 1 diabetes. *J Immunol* 2004. 172:915-922.
- Lehmann PV, Forsthuber T, Miller A, Sercarz EE. Spreading of T-cell autoimmunity to cryptic determinants of an autoantigen. *Nature* 1992. 358(6382):155-157.
- Kaufman DL, Clare-Salzler M, Tian J, Forsthuber T, Ting GS, Robinson P, Atkinson MA, Sercarz EE, Tobin AJ, Lehmann PV. Spontaneous loss of T-cell tolerance to glutamic acid decarboxylase in murine insulin-dependent diabetes. *Nature* 1993. 366(6450):69-72.
- Tisch R, Yang XD, Singer SM, Liblau RS, Fugger L, McDevitt HO. Immune response to glutamic acid decarboxylase correlates with insulitis in non-obese diabetic mice. *Nature* 1993. 366(6450):72-75.
- Herold KC, Bluestone JA, Montag AG, Parihar A, Wiegner A, Gress RE, Hirsch R. Prevention of autoimmune diabetes with nonactivating anti-CD3 monoclonal antibody. *Diabetes* 1992. 41:385-391.
- Jenkins MK, Schwartz RH. Antigen presentation by chemically modified splenocytes induces antigen-specific T cell unresponsiveness in vitro and in vivo. *J Exp Med* 1987. 165:302-319.
- Ledletter JA, Imboden JB, Schieven GL, Grosmaire LS, Rabinovitch T, Lindsten T, Thompson CB, June CH. CD28 ligation in T-cell activation: evidence for two signal transduction pathways. *Blood* 1990. 75:1531-1539.
- Salomon B, Lenschow DJ, Rhee L, Ashourian N, Singh B, Sharpe A, Bluestone JA. B7/CD28 costimulation is essential for the homeostasis of the CD4+CD25+ immunoregulatory T cells that control autoimmune diabetes. *Immunity* 2000. 12(4):431-440.
- Linsley PS, Wallace PM, Johnson J, Gibson MG, Greene J, Ledletter JA, Singh C, Tepper MA. Immunosuppression in vivo by a soluble form of the CTLA-4 T cell activation molecule. *Science* 1992. 257:792-795.
- Lenschow DJ, Zeng Y, Thistethwaite JR, Montag A, Brady W, Gibson MG, Lindsley PS, Bluestone JA. Longterm survival of xenogeneic pancreatic islet grafts induced by CTLA4Ig. *Science* 1992. 257:789-792.
- Salomon B, Bluestone JA. Complexities of CD28/B7: CTLA-4 costimulatory pathways in autoimmunity and transplantation. *Annu Rev Immunol* 2002. 19:225-252.
- 63. Lenschow DJ, Ho SC, Sattar H, Rhee L, Gray G, Nabavi N, Herold KC, Bluestone JA. Differential effects of anti-B7-1 and anti-B7-2 monoclonal antibody treatment on the development of diabetes in the nonobese diabetic mouse. J Exp Med 1995. 181:1145-1155.
- 64. Hathcock KS, Laszlo G, Pucillo C, Linsley P, Hodes RJ. Comparative analysis of B7-1 and B7-2 costimulatory ligands:

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Expression and function. J Exp Med 1994. 180:631-640.

- Ablamunits V, Elias D, Reshef T, Cohen IR. Islet T cells secreting IFN-gamma in NOD mouse diabetes: arrest by p277 peptide treatment. J Autoimmunity 1998. 11:73-81.
- Elias D, Cohen IR. Treatment of autoimmune diabetes and insulitis in NOD mice with heat shock protein 60 peptide. *Diabetes* 1995. 44:1132-1138.
- Elias D, Meilin A, Ablamunits V, Birk OS, Carmi P, Konen-Waisman S, Cohen IR. Hsp60 peptide therapy of NOD mouse diabetes induces a Th2 cytokine burst and downregulates autoimmunity to various beta-cell antigens. *Diabetes* 1997. 46:758-764.
- Bockova J, Elias D, Cohen IR. Treatment of NOD diabetes with a novel peptide of the hsp60 molecule. J Autoimmunity 1997. 10:323-329.
- 69. Raz I, Elias D, Avron A, Tamir M, Metzger M, Cohen IR. beta-cell function in new-onset type 1 diabetes and immunomodulation with a heat-shock protein peptide (DiaPep277): A randomised, double-blind, phase II trial. *Lancet* 2001. 358:1749-1753.
- Chatenoud L, Thervet E, Primo J, Bach JF. Anti-CD3 antibody induces long-term remission of overt autoimmunity in nonobese diabetic mice. *Proc Natl Acad Sci. U S A* 1994. 91:123-127.
- Belghith M, Bluestone JA, Barriot S, Megret J, Bach JF, Chatenoud L. TGF-beta-dependent mechanisms mediate restoration of self-tolerance induced by antibodies to CD3 in overt autoimmune diabetes. *Nature Med* 2003. 9:1202-1208.
- Woodle ES, Hussein S, Bluestone JA. In vivo administration of anti-murine CD3 monoclonal antibody induces selective, long-term anergy in CD8+ T cells. *Transplantation* 1996. 61:798-803.
- Richards J, Auger J, Peace D, Gale D, Michel J, Koons A, Haverty T, Zivin R, Jolliffe L, Bluestone JA. Phase I evaluation of humanized OKT3: Toxicity and immunomodulatory effects of hOKT3gamma4. *Cancer Res* 1999. 59:2096-2101.
- 74. Herold KC, Hagopian W, Auger JA, Poumian-Ruiz E, Taylor L, Donaldson D, Gitelman SE, Harlan DM, Xu D, Zivin RA, Bluestone JA. Anti-CD3 monlconal antibody in new-onset type 1 diabetics mellitus. N Engl J Med 2002. 346:1692-1698.
- 75. Liu E, Moriyama H, Paronen J, Abiru N, Miao D, Yu L, Taylor RM, Eisenbarth GS. Nondepleting anti-CD4 monoclonal antibody prevents diabetes and blocks induction of insulin autoantibodies following insulin peptide B9-23 immunization in the NOD mouse. J Autoimmunity 2003. 21:213-219.

- 76. Phillips JM, Harach SZ, Parish NM, Fehervari Z, Haskins K, Cooke A. Nondepleting anti-CD4 has an immediate action on diabetogenic effector cells, halting their destruction of pancreatic beta cells. J Immunol 2000. 165:1949-1955.
- Cooke A, Phillips JM, Parish NM. Tolerogenic strategies to halt or prevent type 1 diabetes. *Nat Immunol* 2001. 2:810-815.
- Springer TA. Traffic signals on endothelium for lymphocyte recirculation and leucocyte emigration. *Annu Rev Physiol* 1995. 57: 827-872.
- Hanninen A, Taylor C, Streeter PR, Stark LS, Sarte JM, Shizuru JA, Simell O, Michie SA. Vascular addressins are induced in islet vessels during insulitis in nonobese diabetic mice and are involved in lymphoid cell binding to islet endothelium. J Clin Invest 1993. 92:2509-2515.
- Baron JL, Reich EP, Visintin I, Janeway CA. The pathogenesis of adoptive murine autoimmune diabetes requires an interaction between alpha 4-integrins and vascular cell adhesion molecule-1. *J Clin Invest* 1994. 93:1700-1708.
- 81. Kommajosyula S, Reddy S, Nitschke K, Kanwar JR, Karanam M, Krissansen GW. Leukocytes infiltrating the pancreatic islets of nonobese diabetic mice are transformed into inactive exiles by combinational anti-cell adhesion therapy. *J Leukoc Biol* 2001. 70:510-517.
- Rosenstein Y, Park JK, Hahn WC, Rosen FS, Bierer BE, Burakoff SJ. CD43, a molecule defective in Wiskott-Aldrich syndrome, binds ICAM-1. *Nature* 1991. 354:233-235.
- Johnson GG, Mikulowska A, Butcher EC, McEvoy LM, Michie SA. Anti-CD43 monoclonal antibody L11 blocks migration of T cells to inflamed pancreatic islets and prevents development of diabetes in nonobese diabetic mice. *J Immunol* 1999. 163:5678-5685.
- Charlton B, Bacelj A, Mandel TE. Administration of silica particles or anti-Lyt2 antibody prevents beta-cell destruction in NOD mice given cyclophosphamide. *Diabetes* 1998. 37:930-935.
- 85. Taki T, Nagata M, Ogawa W, Hatamori N, Hayakawa M, Hari J, Shii K, Baba S, Yokono K. Prevention of cyclophosphamide-induced and spontaneous diabetes in NOD/Shi/Kbe mice by anti-MHC class I Kd monoclonal antibody. *Diabetes* 1991. 40:1203-1209.
- Mesteller EL, Warner MR, Ferlin W, Judkowski V, Wilson D, Glaichenhous N, Bluestone JA. Peptide MHC class II dimmers as therapeutics to modulate antigen-specific T cell response in autoimmune diabetes. *J Immunol* 2003. 171:5587-5595.